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Animal Tissue Culture SQG 3242 Basics of Cell Culture Media

&

its Supplements

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- Cell Culture: "The process by which either prokaryotic or eukaryotic cells are grown under controlled conditions"
- Mainly implies to the process of culture of animal cells in vitro.
- Important tool in Cell & Molecular biology.
- Cell Based Manufacturing of various products: Vaccines, therapeutic proteins, hormones etc.
- Medium: One of the main parameters to be considered while growing cells *in vitro*
- Major factor affecting process after the Cell line.
- Various defined basal mediums have been developed in last 30 years.
- Initially Balanced salt solutions were widely used.
- Gradually replaced with various vitamins, amino acids, lipids and trace elements.
- Precise media formulations have often been derived by optimizing the concentrations of each ingredient.



Medium

- The choice of medium is empirical
- Defined Medium: Structure and concentration of every component is known
- Undefined Medium: Contain one or more unidentified components.
- Main responses: Measure the growth , cloning efficiency and expression of specific properties i.e production of desired products
- Presterilized stable solutions may be autoclaved at 121°C for 20 min.
- Labile solutions must be filtered through 0.2 um porosity membrane filter.
- Complete Medium contain all necessary defined constituents & supplements

| Medium 199 | Morgan et.al., 1950 |
|------------|---------------------|
| MEM | Eagle 1959 |
| CMRL 1066 | Parker et.al., 1957 |
| DMEM | Dulbecco 1959 |
| Ham's F-12 | Ham 1965 |
| RPMI 1640 | Moore et.al., 1967 |





| Component | Eagle's MEM | Dulbecco's modi- fication | Ham's F12 | CMRL 1066 | RPMI 1 1640 | Component | Eagle's MEM | Dulbecco's modi- fication | Ham's F12 | CMRL 1066 | RPMI 1640 | Component f |
|-------------------------------------|----------------|---------------------------------|--------------|--------------|----------------|---|----------------|---------------------------------|--------------|--------------|--------------|---|
| - | | | | | | L-tyrosine. 2Na | - | - | - | - | -) | Vitamin A acetate |
| mino acids | | - | 8.90 | 25.0 | - 2 | L-valine | 47.0 | 94.0 | 11.7 | 25.0 | 20.0 | Riboflavin |
| L-arginine (free | | • | - | | 200 | Vitamins L-ascorbic acid | : | • | | 50.0 | . } | PO ₄ .2Na Thiamin mono PO ₄ ,2H ₂ O |
| -arginine-HCl | 126 | 84.0 | 211 | 70.0 | - 7 | | | | | 0.010 | | Inorganic salts |
| L-asparagine | | | | | 50.0 | Biotin | - | | 0.0073 | 0.010 | 0.200 | CaCl ₂ (anhyd.) |
| L-asparagine- | - | -15.0 | | - | | D-Ca- | 1.00 | 4.00 | 0.480 | 0.010 | 0.250 | CaCl ₂ .2H ₂ O |
| H ₂ O | | | 20.0 | 20.0 | 30.0 | pantothenate | | | | | | Fe(NO3)3.9H2O |
| L-aspartic acid | - | -13.3 | 30.0 | 20.0 | 50.0 | Calciferol | · · . | • | - | - | - | KCl |
| L-cysteine (free base) | - | | - | - | - | Choline chloride | 1.00 | 4.00 | 14.0 | 0.500 | 3.00 | KH2PO4 |
| L-cysteine | 24 | 48.0 | • | 20.0 | 50.0 | | | | | | | MgCl ₂ .6H ₂ O |
| L-cysteine. 2Na | - | - | | | | Folic acid | 1.00 | 4.00 | 1.30 | 0.010 | 1.00 | MgSO ₄ .7H ₂ O |
| L-cysteine. HCL.H ₂ O | - | - | 35.1 | 260 | - (| i-inositol | 2.00 | 7.20 | 18.0 | 0.050 | 35.0 | NaCl |
| L-glutamic acid | | -14.7 | 75.0 | 20.0 | 66.8 | | 1.00 4.0 | 4 00 | 0.04 | 0.025 | 1.00 | NaHCO ₃ |
| | 584 | 146 | 100 | 300 | 100 4 | Nicotinamide | | 4.00 | .00 0.04 | | | Na2H2PO4.H2O |
| L-glutamine 292 Glycine | - | 30.0 | 7.50 | 50.0 | 10.0 | Pyridoxal. HCl | 1.00 | 4.00 | 0.062 | 0.025 | | Na ₂ HPO ₄ (anhyd) |
| L-histidine (free | | - | - | - | 15.0 | Riboflavin | 0.10 | 0 0.40 | 0.038 0 | 0.010 | 0.20 | Na2HPO4.7H2O |
| base) | | | | 20.0 | 1.4 | | 0.10 | | | 0.010 | | CuSO ₄ .5H ₂ O |
| L-histidine | 42.0 | 42.0 | 21.0 | 20.0 | | Thiamin. HCl | 1.00 | 4.00 | 0.34 | 0.010 | 1.00 | |
| HCI.H ₂ O | | 2 | | 10.0 | 20.0 | | | | | | | FeSO4.7H2O |
| L-hydroxyproline | - | 105 | 3.94 | 20.0 | 50.0 | Vitamin B ₁₂ | - | | 1.36 | | 0.005 | ZnSO ₄ .7H ₂ O |
| L-isoleucine | 52.0 | | 13.1 | 60.0 | 50.0 | Pyridoxine HCl | - | | 0.062 | 0.025 | 1.00 | CaNO ₃ .4H ₂ O |
| L-leucine . | 52.0 | 105 | | 70.0 | 40.0 | | 1 | | | | | Other components |
| L-lysine. HCl | 73.1 | 146 | 36.5 | | | Cholesterol | - | - | - | 0.200 | - | D-glucose |
| L-methionine | 15.0 | | 4.48 | 15.0 | | Para-amino benzoic acid | - | - | - | 0.050 | 1.00 | D-galactose |
| L-phenylalanine | 33.0 | 66.0 | 4.96 | 25.0 | | Nicotinic acid | | | | | | |
| L-proline | - | | 34.5 | 40.0 | | Alcounic acid | | - | - | - | | Lipoic acid |
| L-serine | - | 42.0 | | 25.0 | | Menaphthone | | | | - | | Phenol red |
| L-threonine | 48.0 | 95.0 | 11.9 | 30.0 | | sodium bisulphite | | | | | | Sodium pyruvate |
| L-tryptophan | 10.0 | 16.0 | 2.04 | 10.0 | 5.0 | 3H ₂ O | | | | | | Hypoxanthine |
| L-tyrosine | 36.0 | 72.0 | 5.40 | 40.0 | 20.0 | DI-Â tocopherol PO ₄ .2Na | | - | - | - | -) | Linoleic acid |

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Culture media

- · Provides all the essential nutrients
 - Amino acids, energy substrates, vitamins, minerals, salts, etc
- Maintain constant pH
 - Hepes vs CO₂/HCO₃ buffering
 - Phenol red indicator (yellow-orangemaroon) (acid>>>>>alkaline)



OPENCOURSEWARE Basic Components of Culture Medium

| Nutrient | Substances |
|--------------------|---------------------|
| Water | Energy Sources |
| Nitrogen Sources | Vitamins |
| Bulk Ions | Trace Elements |
| Lipids | Metabolites |
| Non Nutrie | ent Substances |
| Antibiotics | Buffers |
| Protective Agents | Anti Oxidants |
| Metabolites | High M.W Substances |
| 000000000000000000 | |

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Physicochemical Properties - Affectingum

Medium

- **1. pH** for normal cells 7.4, for transformed cells 7.0 7.4
 - phenol red

2. Buffering -

- Commonly used systems: CO₂ ,
- Bicarbonate/HEPES,

3. Oxygen - dissolved oxygen

- Correct O₂ tension so as to meet the requirement and avoid toxicity
- Requirements depends upon type of culture
- Selenium guard against O_2 toxicity

4. Osmolality - 280-310 mOsm/kg

- Measured using depression of the freezing point or elevation of the vapor pressure.
- Helps guard against errors of weighing, dilution.
- Addition of strong acids, bases like HEPES significantly affect.

5. Temperature

- Keep at 4°C-Not use
- Warm at 37C –before use to culture the cells





Animal Sera

- Provides various hormones/ growth factors to stimulate cell proliferation and function
- Less well defined than Serum-free culture systems
- Often difficult to culture cells without serum, but may interfere with studies of specific factors (e.g. insulin, glucose)





Serum Containing Medium

- Traditional undefined medium
- Major source of various nutrients such as growth factors, adhesion factors, minerals, lipids, trace elements etc.
- Commonly used sera: Calf, Fetal bovine, adult horse & human sera
- Horse sera: less metabolism of polyamines, more consistent batch to batch
- Human sera: only used for few cell lines, screening for HIV, hepatitis B virus.
- Promotes cell proliferation, adhesion factors, antitrypsin activity and cell attachment, source of various nutrients





- **Protein Contents:** Albumin carrier of lipids & minerals.
- Fetuin & Fibronectin: promote attachment
- a2 macroglobulin: inhibits trypsin
- Transferrin: makes Iron less toxic and bioavailable.
- **Growth Factors:** main role is in growth stimulation & are mitogenic E.g. PDGF, FGF, EGF, VEGF, Angiogenin etc.
- PDGF major growth factor
- Hormones: Insulin: promotes uptake of glucose & amino acids, mitogenic when bound to IGF – I receptor
- IGF 1/2: mitogenic and stimulate growth
- Hydrocortisone: promote cell attachment
- Minerals: Iron, Copper, Zinc, Selenium essential trace elements required for cell growth. Selenium: important role in detoxifying free radicals by promoting glutathione synthesis.
- **Inhibitors:** TGF β, hydrocortisone cytostatic.
- Apart from these, Carbohydrates (1.0 2.0 mg/mL), various vitamins (10ng -10µg/mL), amino acids are also present.





- Provides various components
- Modulates physiological properties of medium
- Protease inhibitors
- **Provides nutrients** not present in basal medium
- Carrier proteins for low molecular weight substances (e.g. transferrin)
- Help in solubilization of poorly dissolved substances (e.g. apolipoprotein)
- Cell substrate attachment (fibronectin, vironectin)
- Various enzymes
- Proteins which prevent non specific adsorption (e.g. albumin)
- Neutralization of detergents
- Prevents essential nutrients e.g. fatty acids



- i. Potential introduction of animal viruses
- ii. Antibodies against viruses, to which host cell is exposed.
- iii. Availability of high quality
- iv. Undesirable contaminants
- v. High running costs & capital requirements
- vi. Shelf Life & Storage always purchased in bulk
- vii. Physiological variability & consistency
- viii. Downstream Processing
- ix. Characterization of final product laborious.





Media changes

- The purpose of media changes is to replenish nutrients and avoid the build up of potentially harmful metabolic byproducts and dead cells.
- In the case of suspension cultures, cells can be separated from the media by centrifugation and resuspended in fresh media.
- •
- In the case of adherent cultures, the media can be removed directly by aspiration and replaced.





- Some cells naturally live without attaching to a surface, such as cells that exist in the bloodstream.
- Others require a surface, such as most cells derived from solid tissues.
- Cells grown unattached to a surface are referred to as suspension cultures.
- Other adherent cultures cells can be grown on tissue culture plastic, which may be coated with extracellular matrix components to increase its adhesion properties and provide other signals needed for growth



Phosphate Buffered Saline - Ca²⁺ Mg²⁺ Free (PBS)

• Used to wash/remove excess serum that inhibits the function of Trypsin-EDTA.

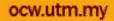
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 Must be warmed in the water bath before use so cells are not shocked by cold liquid.

Trypsin EDTA

- An enzyme used to detach the cells from a culture dish.
- Trypsin cleaves peptide bonds (LYS or ARG) in fibronectin of the extracellular matrix.
- EDTA chelates calcium ions in the media that would normally inhibit trypsin.
- Trypsin will self digest and become ineffective if left in water bath more than 20 minutes.
- Trypsinizing cells too long will reduce cell viability









Water

Quality

Types of Contaminants:

- 1. Inorganics: heavy metals, iron, calcium, chlorine
- 2. Organics: by products of plant decay, detergents
- 3. Bacteria: Endotoxin or pyrogens
- 4. Particles: Colloids or particles

Purification methods:

- a. Distillation
- b. Deionization
- c. Reverse osmosis
- d. Ultra Filtration





- Most common: *glucose* Others: *Fructose, galactose, mannose, maltose* etc
- D-Glucose: main source of energy
- Metabolized by glycolysis and energy is released in form of NADH and ATP.
- Also metabolized via PPP pathway where NADPH is released.
- Stored in cells as glucose-6-phosphate/glycogen
- Cannot penetrate cell membrane, require help from transporter proteins
- Either by osmosis or by energy utilization in form of ATP.
- Mainly transferred via Glut I transporter
- 1 10g/L
- Higher concentration creates negative effect

High level glucose = high level of lactate = low pH = detrimental to cells = requirement of base = increase in osmolality

• Lead to glucose related oxidative and carbonyl stress.





_ – Glutamine (1–20mM)

- Major Carbon Source in case of low glucose levels & Nitrogen source
- Support cell growth and amino acid uptake
- Metabolic hydrolysis to glutamic acid
- Spontaneously decomposes, overproduction of ammonia
- In vivo ammonia produced is converted to urea
- Invitro ammonia produced due to glutamine degradation accumulates
- At physiological pH, inorganic ammonia converted to organic nitrogen either in the form of amine of glutamate or amide of glutamine which act as primary reservoir for nitrogen.
- Acts as precursor of glutamate role in transamination
- Most commercial medias free from glutamine included either in basal formulation or in liquid formulation at time of use.





Exercise

• At what temperature serum and tyrpsin can be stored ?

• Why?





- Basal Salt Mixtures mainly containingCaCl₂, MgCl₂, KCl₂, Potassium Phosphate Monobasic (Anhydrous), NaCl₂, Sodium Phosphate (Dibasic)
- Provide bulk ions for cell metabolism e.g Na⁺, K⁺
- Osmotic balance maintenance & provide buffering system
- Cofactors in various enzymatic reactions
- Cell adhesion (Ca^{2+,} Mg²⁺⁾
- Binding of Iron to transferrin (HCO₃⁻)
- Contribute to Osmolality of medium (280-310 mOsm/kg)

Trace Elements

Iron, Manganese, Zinc, Molybdenum, Selenium, Vanadium, Copper

Lipids & Phospholipids Precursors

- Cholesterol, Fatty acids
- Phospholipids phosphotidyl choline, phosphotidyl ethnaolamine, sphingomyelin
- Choline, Inositol, Ethanolamine.





Antibiotics: more inhibitory to bacteria then to cell culture cells

- Commonly used antibiotics: Gentamycin ,Fungizone Penicillin streptomycin
- Cautious selection
- **Buffers:** Sodium Bicarbonate 44mM in DMEM, 12mM in F12, 26mM in circulatory blood.
- 5-10% CO₂ required as media containing bicarbonate becomes alkaline very rapidly due to loss of CO₂
- Low pKa
- Alternate buffers: Sodium beta glycerophosphate, HEPES osmolality
- Phenol Red: pH indicator, might be estrogenic

Protective Agents: Pluronic Surfactants – Polyoxyethylene group

- Toxicity effect.
- ▶ 0.01 0.1% working concentration.
- Commonly used pluronics: F68, F88, F77





Antioxidants

- Superoxide radicals & Hydrogen Peroxide harmful
- Generated during normal respiratory metabolism by xanthine oxidase/photo oxidation of riboflavin,tryptophan.
- Commonly used Vitamin E, Taurine, Bilirubin, transferrin, various amino acids, selenium, catalase, glutathione.

Metabolites & Conditioning Factors

- Produced by cells usually of 2 types:
 - (i) Growth Inhibitory: Lactate, ammonia, CO2, TGF beta etc
 - (ii) Growth Stimulatory: TGF alpha, IGF 1, IL 2, PDGF





- 2. Insulin Like Growth Factor one order less as compared to insulin
- Can replace insulin, bind to IGF-IR
- IGF-1 affects protein & carbohydrate metabolism
- Also stimulates glycolysis and protein synthesis
- Increase DNA synthesis, affect cell proliferation & differentiation
- 3. Transferrin 1-30µg/ml
- 80KDa with N/C terminal iron binding domains
- Binds with high dissociation constant
- Can be replaced with hemoglobin, ferrous sulfate etc





- Mainly divided as essential & nonessential
- Wide range of optimal concentration and play important role in primary protein structure
- Semi log plot of growth response vs. nutrient scale
- Initially only essential amino acids but now both are supplied.
- Amino acid hydrolysates an important substitute for serum, can be autoclaved but are undefined. E.g bactopeptone, tryptose etc.
- Some important amino acids are:
- Proline: required by mutant CHO cells
- Serine: required for high clonal densities
- Aspargine: required mainly by tumor producing cells
- L-cysteine: active role in protein synthesis; important in glutathione synthesis thereby prevent cells from oxidative stress
- Glycine: mainly used with methotrexate and aminopterin which leads to folic acid deficiency.



Inspir

- Eagle's MEM contains only water soluble vitamins, rest derived from serum.
- Affect cell survival & growth rates.
- Usually added empirically till the actual effect determined

| Ascorbic acid | Synthesis of collagen |
|--------------------------------|---|
| Vitamin A | Growth & differentiation of some cells |
| Vitamin K | Many Vitamin K dependent proteins |
| Vitamin E | Antioxidant |
| Vitamin D | Regulation of Ca ²⁺ , important to maintain at appropriate levels. |
| Thiamine pyrophosphate | Catalyst in the transfer of carboxyl group, transaldolases, transketolases |
| Pyridoxal phosphate | Catalyses transamination |
| Biotin | Carrier of CO ₂ , required for functioning of pyruvate dehydrogenase, pyruvate carboxyase & fatty acid synthesis |
| Vitamin B12 | Important for methylation, degradation of ami acids & fattyacids |
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Summary

- One of the major factors considered in cell culture is the choice of Medium
- Type of Medium & its constituents affect culturing & passage of cells.
- Till date, most of the selection is done Empirically.
- Depending upon the previous data obtained & medium used for different cell lines.
- Selection of energy & nitrogen source plays a critical role along with its concentration.
- Adaptation of a cell line from serum containing to serum free medium is carried out using serial subcultures by gradually reducing the serum concentration.
- Concentration & effect of the amino acids used should be priorly optimized.

Conclusion:

Although relative simplicity of retaining serum in medium is lucrative, requirements of governing bodies & various disadvantages of serum have lead to the development of serum free compositions. The main intention being to simplify the process of cell culturing, protein purification and to eliminate all potential sources of infection from the medium.



Phenol red indicator

• Alkaline?

– Colour and pH

- Acidic?
 - Colour and pH





Contamination

- Normally:
 - Bacteria
 - -Yeast
 - -Virus
 - Pleuropneumonia group
 - Protozoa *





Detection

Inverted microscopic

- Bacteria
 - Can be detected immediately
 - Takes about 24-48 hours
 - Easily found using the microscope/ naked eyes??
- Yeast
 - Slow growing
 - Takes few days to be detected using the microscope
 - Worst- naked eyes
- Virus
 - Difficult to traces using the microscope. Size smaller than bacteria
 - Can takes ages to be detected.
 - How we know there are virus in the cell media?





- Cells stressed
- Media colour change quickly-cells still not confluent
- Debris occurs
- Pleuropneumonia group
 - Known as PPLo/mycoplasma
 - Serious problem of chronic contamination
 - Present in many animal tissue
- Protozoa
 - Contaminating the fresh isolated tissues
 - Easily recognised on microscopic examination





Sterility Test

- To find out which chemicals being infected
 - Supplemented media --incubator
 - Supplemented media+trypsin-incubator
 - Supplemented media +PBS-incubator
 - Check everyday for a week
- Or
 - Spread about 1ml of supplemented media/tyrpin/PBS on agar plate
 - Leave in incubator –check for microorganism growth



Elimination of contamination

- Do not rescue the contaminated cell lines
- Discard any contaminated cells (flasks, plates)-autoclave ASAP
- Check the sterility of the chemicals
- Incubator cleaning, biohazard/laminar flow cleaning, change/wash lab coat, change water bath water /+ waterbath treatment water







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